

Evaluation of Antimicrobial Activity of β -tricalcium Phosphate/Calcium Sulfate Mixed-up with Gentamicin: In-Vitro Study

Research Article

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Abstract

Objectives: An in-vitro study was designed to investigate the antimicrobial activity of β -tricalcium phosphate/calcium sulfate in comparison to Xenograft, blended with either gentamicin or saline.

Methods and materials: Employing Kerby-Bauer disk diffusion test, twelve Petri dishes (90 mm) with two models of culture mediums were used to cultivate three bacterial strains: Muller-Hinton Agar (for *P. Aeruginosa*/*S. Aureus*) and Sheep Blood Agar (for *Enterococcus faecalis*). Then, studied materials were divided into four groups (Bovine, Bovine with gentamicin, EthOss and EthOss with gentamicin), mixed-up and transmitted into their holes in each dish. After 24h of incubation, inhibition zones were noted and measured by a digital caliper. In the end statistical analysis were completed with One Way ANOVA followed by Tuckey HSD test on SPSS17.

Results: One Way ANOVA and Tuckey HSD remarked a statistically significant difference between all pairs for *Enterococcus faecalis* ($p < 0.05$), except of Bovine/ EthOss pair. When it comes up to *Staphylococcus aureus*, a significant difference was observed between all pairs ($p < 0.05$), except Bovine/ EthOss and EthOss Gentamicin/ Bovine Gentamicin pairs, both were effective in bacterial elimination. Finally, only Bovine Gentamicin was functional with *Pseudomonas aeruginosa* ($p < 0.05$).

Conclusion: β -tricalcium phosphate/calcium sulfate and Xenograft antibacterial abilities were improved when mixed with Gentamicin. Xenograft was preferable as an antibiotic carrier when it comes to *Pseudomonas aeruginosa*.

Keywords: β -tricalcium Phosphate; Calcium Sulfate; Xenograft; Gentamicin; Antimicrobial.

Introduction

Dental implantation is a common treatment for tooth loss, despite survival rates of 96.33% after 8 years of the procedure, biological complications are likely to occur, including peri-implantitis, which includes bone loss and peri-implant mucositis.[1]

The principle goal of peri-implantitis treatment is to manage the infection, prevent further bone resorption, maintain aesthetics and enhance bone regeneration in the area of bone loss.[2]

Various types of bone grafts have been used for bone regeneration. Bone grafts are classified according to their source to: Autogenous Bone Grafts (tibia, Iliac, ramus), Allograft (FDBA,

DFDBA), Xenograft (Bovine) and Alloplastic Bone Graft (Calcium Sulfate, Calcium Phosphate).[3]

Peri-implantitis etiology is similar to periodontitis. In both of them germs attach to the implant's or tooth's surface causing bone resorption.[2]

The main goal of treatment is the removal of bacterial plaque.[2] *Pseudomonas aeruginosa* (a Gram-negative, facultative anaerobic rod bacterium), *Staphylococcus aureus* (Gram-positive, facultative aerobic, round-shaped, bacterium) and *Enterococcus faecalis* (Gram-positive, facultative anaerobe) were frequently found in peri-implantitis. In addition to that, they were accused with implant failure.[4, 5]

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Some researchers have suggested adding antibiotic to the bone graft that used for regenerative therapy around Implants to reduce the bacterial Load. [6]

The benefits of local release is the ability to provide high local antibiotic concentrations without systemic toxicity. It has been suggested that such high concentrations can even penetrate a bio-film.[7, 8]

One of the most used material is Polymethylmethacrylate (PMMA). This material is an acrylic non-absorbable material that must be removed in a surgical procedure. PMMA beads may continue to excrete low levels of antibiotic even up to five years, which may generate multi-drug resistant organisms. Furthermore, once the antibiotic levels are too low to kill organisms the PMMA itself can become colonized. [9, 10]

Calcium sulfate bone graft is an absorbable material does not need a surgical procedure to remove, several studies have been aimed to combination antibiotic with calcium sulfate substitutes depending on it biocompatibility, porosity, and biodegradability. [10]

Gentamicin is aminoglycoside, which is used for treatment of serious infections caused by aerobic gram-negative bacilli such as *Pseudomonas aeruginosa*. It has a limited effect on gram-positive germs like *Staphylococcus aureus* and *Enterococcus faecalis*. Yet, Gentamicin's clinical utility is limited due to its serious toxicity. [11] Coating titanium implants with Gentamicin found no negative impact on osteoblast function. [12]

The aim of this study is to evaluate the efficacy of the biphasic absorbable bone graft (35% calcium sulfate/65% β -tricalcium phosphate) mixed with either normal saline or gentamicin 2 mg/ml, in comparison to bovine bone graft mixed with either normal saline or gentamicin 2 mg/ml in inhibiting *P.aeruginosa*, *S.aureus* and *E.faecalis* growth.

Methods and Materials

Three clinical bacterial strains isolates were collected from different patients who were administrated to Al-Mowasat Hospital- apartment of Bacteriology tests Laboratory. The study was accomplished with diffusion test method. Twelve Petridishes (90 mm) with two models of bacterial culture mediums were used (Muller-Hinton Agar for *P. Aeruginosa* and *S. Aureus* / Blood Agar for *Enterococcus faecalis* isolated from necrotic root canals). To perform Kirby Bauer disk, bacterial density was controlled by PhonexSpec at 0.5 McFarland Standard (1.5×10^8 CFU/ml). Then a sterile swabs was dipped into inoculum tube, to remove the ex-

cess fluid the swab was pressured around the tube walls. Bacteria was inoculated over its specified agar dish, then was left at 37° for 20 minutes to dry.

At the margins of each agar dish a hole of (4mm) depth and (6mm) diameter was punched; the diameter was defined to simulate the standard diameter of the antibiotic sensitivity disk. Four mixed materials were prepared:

1. Biphasic absorbable alloplastic bone substitute (EthOss®, EthOss Regeneration Ltd., Silsden, UK) which consists of β Tricalcium Phosphate (65%) and Calcium Sulfate (35%) mixed with normal saline.
2. EthOss mixed with Gentamicin 2mg/ml (Gentacine®, Ibn Hayyan Pharma, Homs, Syria).
3. Bovine bone graft (MedPark Bone-D®, MedPark, Busan, Korea) mixed with normal saline.
4. MedPark Bone-D mixed with Gentamicin 2mg/ml.

Then each mix was freshly transformed to fill its hole in each agar dish, then we kept it for one hour at room temperature to insure the expansion of the materials through the Agar. Thereafter, they were incubated at 37°C for 24 hours before inducing the test.

Inhibition zones surround each hole were measured by the digital caliper, to observed the anti-bacterial activity (higher inhibition zone diameters) of the tested mixture.

Finally, SPSS program was used to accomplish the statistical descriptive and analytic processes. At significance level of 0.05, multiple comparison between groups was done using One Way ANOVA test followed by Tuckey HSD.

Results

Statistical analysis were performed with SPSS software. Means, standard deviations, minimum and maximum values of each tested material according to its efficacy in inhibiting microbial growth of tested microorganisms is described in table 1.

Shapiro-Wilk normality test stated a normal distribution of the values. Therefore, One Way ANOVA test was carried out, it conducted a significant difference for all studied materials against each studied microorganisms ($p < 0.05$) table 1.

To compare between material pairs Tuckey HSD test was accomplished. A significant difference was seen between all pairs for *Enterococcus faecalis* ($p < 0.05$), except Bovine/EthOss pair. For *Staphylococcus aureus*, a significant difference was observed between all pairs ($p < 0.05$), except Bovine/EthOss and EthOss

Figure 1: a. b. measure the diameter of inhibition zone by digital caliper.



Table 1.

Microorganisms	Tested material	Mean	Std. Deviation	Minimum	Maximum	ANOVA Sig.
Enterococcus faecalis	Bovine	0	0	0	0	0
	EthOss	0	0	0	0	
	Bovine Gentamicin	30.565	0.575	29.9	31.2	
	EthOss Gentamicin	27.307	0.229	27.1	27.6	
Staphylococcus aureus	Bovine	0	0	0	0	0
	EthOss	18.178	8.116	10.7	26.2	
	Bovine Gentamicin	37.715	2.408	36.2	41.3	
	EthOss Gentamicin	34.51	0.336	34.2	34.8	
Pseudomonas aeruginosa	Bovine	0	0	0	0	0
	EthOss	0	0	0	0	
	Bovine Gentamicin	20.838	1.223	19.4	22.3	
	EthOss Gentamicin	0	0	0	0	

Table 2.

Multiple Comparisons Tukey HSD	(I) Group	(J) Group	Sig.	
Enterococcus faecalis	Bovine	EthOss	1.000	
		Bovine Gentamicin	.000	
		EthOss Gentamicin	.000	
	EthOss	Bovine Gentamicin	.000	
		EthOss Gentamicin	.000	
	Bovine Gentamicin	EthOss Gentamicin	.000	
	Staphylococcus aureus	Bovine	EthOss	1.000
			Bovine Gentamicin	.000
			EthOss Gentamicin	.000
EthOss		Bovine Gentamicin	.000	
		EthOss Gentamicin	.001	
Bovine Gentamicin		EthOss Gentamicin	.713	
Pseudomonas aeruginosa		Bovine	EthOss	1.000
			Bovine Gentamicin	.000
			EthOss Gentamicin	1.000
	EthOss	Bovine Gentamicin	.000	
		EthOss Gentamicin	1.000	
	Bovine Gentamicin	EthOss Gentamicin	.000	

Gentamicin/Bovine Gentamicin pairs. Finally, only Bovine Gentamicin revealed a significant difference with all compared materials for Pseudomonas aeruginosa (p<0.05) table 2.

Discussion

Successful perimplantitis treatment requires effective elimination of persistent bacterial population from implantation region.[13] This is carried out through mechanical and chemical treatment of implant surface.[14] In addition to that, bone graft might possess some antimicrobial activity, driving the managed perimplantitis area to promising long-term outcomes.

Staphylococcus aureus, Enterococcus faecalis and Pseudomonas

aeruginosa were tested as long as they frequently exist in implant-related bone infections.[15, 16]

Agar diffusion method is recognized as standardized method to determine primary antimicrobial capacity of the studied materials. However, it has some limitation such as its inability in recognizing whether the studied material has bactericidal or bacteriostatic effects, therefore those results must be read with caution.

Group 1 (Enterococcus faecalis):

Both bovine and EthOss beads did not have any antimicrobial activity. Bovine is chemically inactive substance without any antimicrobial ability. When it comes to β-tricalcium phosphate/calcium

Figure 2. a. The four materials after transformed to its holes in blood agar.
b. The inhibition zone after 24h.

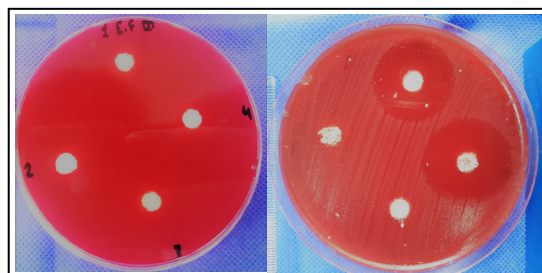


Figure 3. a. The four materials after transformed to its holes in Muller-hinton agar.
b. The inhibition zone after 24h.

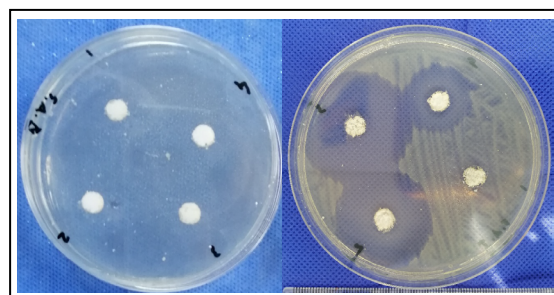
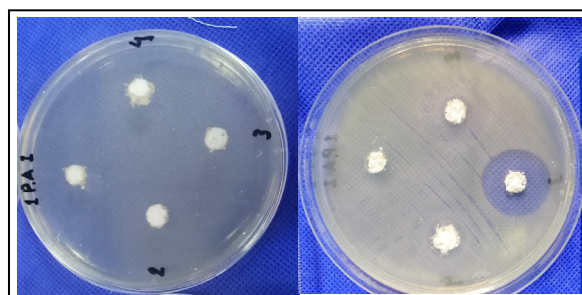


Figure 4. a. The four materials after transformed to its holes in Muller-hinton agar.
b. The inhibition zone after 24h.



sulfate, there was no previous studies on its antimicrobial activity for *E.faecalis*.

On the other hand, mixing the studied bone-grafts with gentamicin showed a significant antimicrobial action; it seems that they acted as an antibiotic carrier helping in elimination of bacterial load. However, it was obvious that bovine with gentamicin had a slightly larger inhibition zone than EthOss with gentamicin (30.56 ± 0.57 mm, 27.30 ± 0.22 mm); it can be explained by the reduction of gentamicin's efficacy at acidic media [17], it was mentioned by Fergusson et al. that EthOss transforms from neutral to an acidic media at dissolve-stage.[6]

Group 2 (Staphylococcus aureus):

Bovine beads did not have any antimicrobial activity. However, EthOss demonstrated significant antimicrobial activity with an inhibition zone of 18.17 ± 8.11 mm diameter. Shizou et al studied calcium phosphate against *S.aureus* implanted in Muller-Hinton agar, they conducted that an inhibition zone of 32.2 ± 2.5 mm diameter was formed. [18]

Adding gentamicin to the studied bone-grafts displayed a significant antimicrobial action.

Bovine with gentamicin had a slightly larger inhibition zone than

EthOss with gentamicin (37.71 ± 2.40 mm, 34.51 ± 0.33 mm), without any statistical difference between them.

Group 3 (Pseudomonas aeruginosa):

When it comes to *P.aeruginosa* inhibition abilities; bovine, EthOss and EthOss with gentamicin antimicrobial activities were none. The acidic microenvironment produced by EthOss at dissolve stage may have affected gentamicin's efficacy. [17] Gentamicin's concentration after being mixed with EthOss seems to be lower than the Minimum Inhibitory Concentration (MIC) needed to kill *P.aeruginosa*. Which requires more investigation in further studies.

Bovine with gentamicin showed an inhibition zone of 20.83 ± 1.22 mm diameter, it played the role of a physical carrier for gentamicin without any interactions in-between.

In present work, EthOss with gentamicin exhibited antimicrobial activity against all previous strains except *P.aeruginosa*. On the contrary, Bovine with gentamicin was effective on all strains so it might be better in immediate implantation cases that have a periapical lesion or in apicoectomy procedures.

EthOss solitarily or with gentamicin can be used in perimplantitis surgical regenerative interventions, due to its antimicrobial effect

on *S.aureus*, this strain is frequently associated with implant-related bone infections [16].

Agar diffusion test standards of inhibition zone for Gentamicin-measures: 19-27mm for *S.aureus*, 16-22mm for *P.aeruginosa*, and 15-25 mm for *E. faecalis*. [21, 22]

When it comes to *E.faecalis* group, inhibition zones diameter of Bovine with gentamicin and EthOss with gentamicin were 30.56 ± 0.57 mm and 27.30 ± 0.22 mm respectively. Moreover, *S.aureus* group diameters for EthOss, Bovine with gentamicin and EthOss with gentamicin were 18.17 ± 8.11 , 37.71 ± 2.40 mm and 34.51 ± 0.33 mm respectively. Considering these results, and comparing them to the critical values of gentamicin, it can be concluded that studied bone grafts preserve the antimicrobial action of gentamicin playing the role of appropriate medication carrier to the cured area. On the other hand, in *P.aeruginosa* group only Bovine with gentamicin had an inhibition zone with diameter of 20.83 ± 1.22 , indicating an antimicrobial effect in comparison with critical gentamicin values.

Smaller inhibition zone diameter of EthOss with gentamicin compared to Bovine with gentamicin might be referred to the ingress of gentamicin solution into the composition of EthOss while it hardens, decreasing the initial concentration of the medicament.[6]

Conclusion

Last but not least, this in-vitro study reported that mixing β -tricalcium phosphate/calcium sulfate, and Xenograft with Gentamicin enhanced favorable antibacterial abilities to manage infected bone zones. Additionally, Xenograft was preferable as an antibiotic carrier especially when it comes to *Pseudomonas aeruginosa*. Finally, β -tricalcium phosphate/calcium sulfate bone grafts appears to have minimal antimicrobial activity against *staphylococcus aureus*.

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